Regenerative Strategies for Anterior Esthetic Rehabilitation: A Clinical and Histologic Case Report

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Abstract: In order to solve the substantial challenge of achieving natural esthetics in compromised situations, a number of techniques and materials have been developed to restore deficient alveolar bone and maintain existing bone and soft tissue. This article reviews developments in both areas and reports on a clinical treatment in which a new form of xenograft in conjunction with a cross-linked, cell-occlusive membrane was used in a severely compromised anterior maxilla in preparation for placement of platform-switched implants.

With the rapid advancement in understanding of the biomechanical interaction of implants and bone, implants no longer can be deemed successful simply based on their ability to retain a functional restoration. Instead, the faithful recreation of natural esthetics also has become imperative. Nowhere is that more important than in the anterior maxilla.

Numerous factors may inhibit clinicians from placing anterior maxillary implants in such a way as to yield optimal restorative results. Several clinical and histologic studies have documented the dynamic resorptive process that unfolds after tooth extraction.[1-7] The alveolar ridge dimensions may be altered both vertically and horizontally, with up to half the native ridge width lost during the first 12 months and approximately two thirds of that reduction occurring within the first 3 months.[3,8,9] This characteristic loss of osseous volume caused by normal remodeling has major consequences for implant reconstructive dentistry, particularly when encountered in the esthetic zone. The ability to obtain an esthetic result is further complicated when the patient presents with compromised native morphology, pathology, trauma, or developmental anomalies.

In an attempt to solve some of these challenges, two broad trends have been developed within implant dentistry. The first is the proliferation of techniques and materials available for preventing or regenerating deficiencies in crestal height. The second is the growing understanding of biomechanical strategies for maintaining existing hard and soft tissue, most notably by means of the implant design itself.

BONE AUGMENTATION

In horizontally and vertically atrophic ridges, extraction sockets, implant-associated defects, and other anatomic structures that may inhibit proper implant positioning, a number of successful bone regeneration techniques have been well documented. These include ridge splitting and expansion;[10-12] guided bone regeneration (GBR);[13,14] distraction osteogenesis;[15] onlay, inlay, and veneer grafts;[16-18] nerve lateralization;[19] and sinus augmentation.[20,21]

When employing such techniques, any bone regeneration material used should optimally be osteoinductive and osteoconductive, easily obtainable, safe, biocompatible, malleable, space-maintaining, and stable after positioning. Several categories of materials have become acceptable alternatives for filling and regenerating bone defects, including autografts, allografts, alloplasts, and xenografts. However, no material meets all the aforementioned criteria.

Autogenous bone, considered historically to be the gold standard, has osseoinductive, osseoconductive, and osteogenic properties and poses no risk of immunologic rejection. Yet, the harvesting of autogenous bone has been associated with various complications and morbidity,[22-27] and it may be difficult to obtain in sufficient quantity. High resorption rates and limited viability for autogenous block grafts also have

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been reported,[28-31] and survival rates of implants placed in autogenous grafts have not always exceeded those of implants placed in bone regenerated with alternative materials.[32,33]

Allograft bone, derived from cadavers and obtained from tissue banks, is the most popular alternative to autogenous bone. It offers the substantial advantage of allowing the patient to avoid a second surgical procedure, with the attendant risks of pain, complications, and morbidity. Available in various shapes and sizes, allograft bone also has fewer supply restrictions than autograft material. However, the donor, tissue bank, and means by which the allograft is processed can dramatically affect its osseoinductive potential (which can range from none to substantial) and the response of the recipient site to its introduction.[34-36]

Synthetic bone graft materials (alloplasts) constitute a third category of widely used bone-grafting materials. Engineered with properties resembling bone and allowing for bone regeneration, alloplasts include such nonhuman, artificially produced materials as calcium sulfate, calcium phosphate, hydroxyapatite, and bioactive glass. Their advantages consist of their abundant supply, long shelf lives, and lack of potential disease transmission. Excluding a handful of clinical case reports, alloplasts alone have historically displayed evidence of variable clinical successes due to their lack of osteoinductive properties and questionable osseoconductive properties. Their resorption rates may be unacceptably fast or slow, and their capacity to predictably regenerate bone has not yet been demonstrated consistently.

Xenografts constitute the final category of bone-grafting materials in common use. Derived from animal sources, xenografts have been developed largely in response to the donor site complications associated with autografts, as well as the limited supplies of both autogenous and allograft material. Porous bovine-derived material is by far the most common variety of xenograft in use today. Such material has been demonstrated to be highly biocompatible.[37] Although concern has also been raised about the possible transmission of bovine spongiform encephalopathy (BSE) by means of xenografts, current data indicate that the risk of BSE transmission is virtually nil.[38,39] In fact, no case of BSE traced to long bones from bovine sources has been reported.[40]

Many of the advantages associated with xenograft material mirror those of allografts. Using bovine bone eliminates the need for a second surgical site and the possibility of any attendant complications. The material is available in various configurations, in unlimited supply, as well as at lower cost than either autogenous or allogeneic bone. It has been demonstrated

to be an effective regenerative matrix for periodontal defects, sinus floor elevation, and ridge augmentation prior to implant placement. Histologic analysis has found bovine bone particles to be well incorporated within newly regenerated grafted bone. While some human[41] and animal studies[42] have concluded that porous bovine bone mineral lacks both osseoinductive and osseoconductive properties, other researchers have reported a high degree of osseoconductivity.[37,43-49]

For any of the categories of bone-grafting materials, following the principles of GBR and using an appropriate barrier membrane can enhance the likelihood of obtaining optimal results.[13] The ideal properties of a barrier for GBR procedures include the ability to exclude unwanted epithelial cells and maintain a space for osteogenic cells to repopulate the graft site. Nonresorbable membranes, while documented to be effective,[50] require a second surgical entry and thus increase the patient's cost, discomfort, and psychological stress. Other problems associated with nonresorbable membranes include increased tissue trauma and wound healing

Figure 1 The patient presented with a suppurative fistula apical to the maxillary left central incisor.

Figure 2 Radiograph of the maxillary left central incisor displaying periapical radiolucency.

complications, such as membrane exposures, infection, bacterial contamination, and poor regenerative outcomes.[51-53]

Resorbable collagen membranes avoid the drawbacks associated with the need for a second membrane removal surgery and have a number of other benefits. Biocompatible[54,55] and nonimmunogenic,[55] they promote hemostasis and have been shown to have a chemotactic effect on periodontal ligament cells[56,57] and gingival fibroblasts. They promote osteoblast proliferation, as well as increased secretion of transforming growth factor-β1, which is involved in bone remodeling.[58] Non-cross-linked resorbable collagen membranes also often foster a very rapid vascularization and excellent tissue reaction subsequent to the vascularization.[59,60] However, their resorption profile is roughly only 4 to 8 weeks, which is too rapid, given that a statistically significant increase in bone formation can occur in the 8th to 16th week after grafting (in the absence of complications).[61]

To extend the absorption profile of collagen membranes, cross-linking techniques have been developed. Cross-linked membranes, while semipermeable and allowing for an exchange of nutrients, have been demonstrated to exclude epithelium and connective tissue for at least 6 months.[62] This gives them the potential to provide most of the benefits of nonresorbable membranes, without the latter's drawbacks.

BONE PRESERVATION

The most predictable method to achieve a natural appearance prior to implant placement is to preserve the patient's native hard and soft tissue before they are lost. Often, this is not possible; however, the use of minimally traumatic extraction techniques is an important first step in treatment. Another area where vertical bone loss occurs routinely after implant placement is in the peri-implant crestal bone. Bone loss up to the first implant thread has been considered normal since the earliest days of implant dentistry[63] and has not threatened the implant's long-term survival. However, as the hard tissue retreats, the soft tissue subsequently is compromised. This consequential loss of soft tissue has a high likelihood of impacting the overall esthetic result.

Figure 3 Full-thickness reflection revealed a defect of the alveolar facial plate measuring approximately 8 mm to 10 mm circumferentially.

Figure 4 Implant partially encased within native bone and defect.

Figure 5 Defect and residual implant threads augmented with Endobon Xenograft Granules.
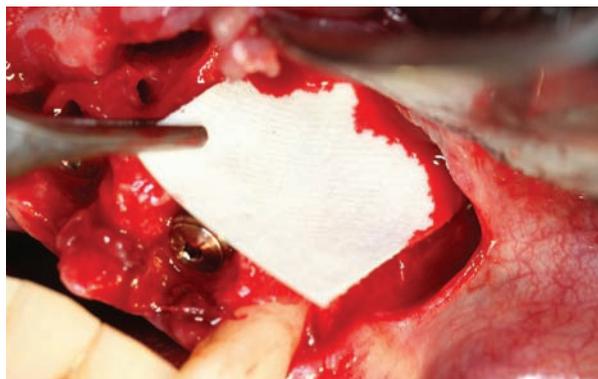

Figure 6 A dry OsseoGuard membrane was inserted under the facial flap to verify the apex of the lesion covered, as well as to assist in soft-tissue reflection.

A number of researchers have theorized the crestal bone loss that typically occurs after connection of a healing abutment is related to development of the biologic width. As bacterial contamination of the implant-abutment junction (IAJ) triggers an inflammatory response, a new biologic width must develop at the IAJ in order to seal off and protect the crestal bone from the oral environment. Thus, the bone recedes to accommodate the peri-implant mucosa.

To forestall this cascade of events, the concept of platform switching has been developed. Platform switching involves the use of a smaller-diameter abutment on a larger-diameter implant collar, a design that shifts the inflammatory cell infiltrate away from the crestal bone. A number of successful long-term clinical cases have been reported, and 1-year postoperative results of clinical research investigations have documented a significant reduction of crestal bone loss around platform-switched implants.[64-66]

The following presentation illustrates a case in which GBR using a new form of xenograft in conjunction with a cross-linked-collagen cell-occlusive membrane was used to restore a severely compromised anterior maxilla before placement of implants with built-in platform switching in the central incisor positions.

CASE REPORT

A healthy 48-year-old male with a moderate-to-low smile line presented with severe pain above his left maxillary central incisor. A review of his social history revealed that he had smoked approximately one package of cigarettes daily for 15 years. The intraoral examination revealed the presence of a fluctuant mass apical to the maxillary left central incisor and an abutment for an existing fixed three-unit bridge (Figure 1). Radiographic findings confirmed a large (8 mm x 10 mm) periapical radiolucent pathology (Figure 2).

The patient was referred for immediate endodontic evaluation; those findings helped determine that the tooth root had a vertical fracture due to an enlarged post. The right maxillary central incisor was also found to be periodontally compromised. On returning to the surgical office, the patient was advised that smoking has been associated with an increased risk of impaired wound healing and implant failure.[67] Nonetheless, he gave informed consent for removal of both maxillary central incisors and immediate reconstruction of the compromised alveolar ridge.

Following administration of local anesthetic (2% lidocaine and epinephrine 1:100,000) and 2 g of prophylactic amoxicillin, impressions were made for on-site fabrication

Figure 7 Two OsseoGuard membranes hydrated with the patient's blood and positioned under the facial and palatal flaps.

Figure 8 At the second-stage surgery, Encode Healing abutments were placed.

Figure 9 Postoperative radiograph confirmed that the alveolar crestal height was maintained.

of a provisional restoration. The two central incisors were extracted with minimal trauma to the alveolar bone and both sockets debrided of granulomatous tissue. However, it was not possible to verify conclusively that all the granulation tissue was successfully excised, so a full-thickness mucoperiosteal flap was reflected from canine to canine. This revealed a bony deficiency at the base of the nasal floor measuring approximately 8 mm to 10 mm circumferentially (Figure 3). Due to the minimally traumatic extraction, the thickness
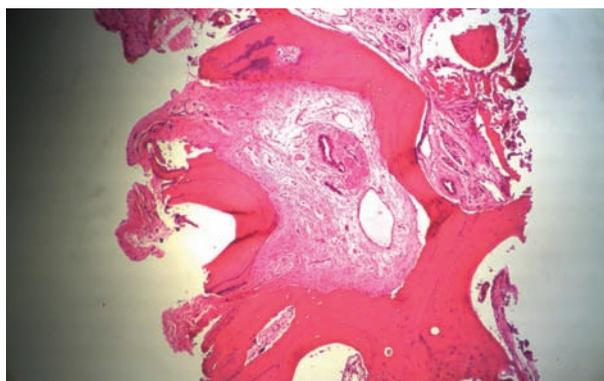
Figure 10 The definitive restoration.

Figure 11 Histologies from regenerated sites at 6 months. Low-power microscopic examination revealed new bone formation in conjunction with a number of unresorbed xenograft particles (hematoxylin and eosin [H&E] stain; original magnification x10).
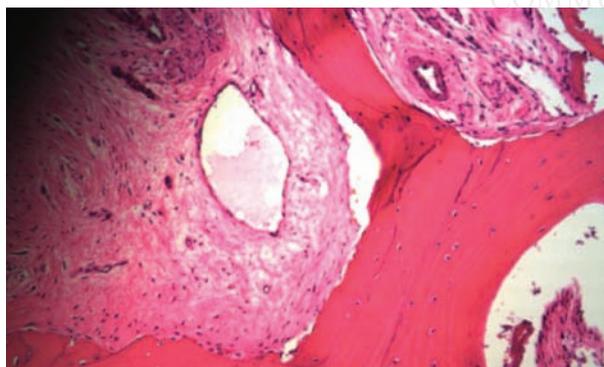

Figure 12 Higher-power microscopic examination showed newly formed bone and medullary spaces with well-embedded xenograft (H&E stain; original magnification x30).

of the marginal crestal alveolus appeared to be adequate for primary stabilization of two expanded platform dental implants.

After removal of all remaining granulation tissue, osteotomies were created under copious irrigation within the extraction sites. Two 4/5/4 mm diameter x 11.5 mm NanoTite™ PREVAIL® Implants (BIOMET *3i*, www.biomet3i.com) were placed, and their stability was verified by torquing both implants to greater than 40 Ncm.

In both sites, the flare of the implant platform effectively obliterated the socket. However, at the left central incisor site, the threads of the implant apex were exposed within the defect (Figure 4). Approximately 0.5 cc of Endobon® Xenograft Granules (BIOMET *3i*) were mixed with sterile saline and allowed to hydrate for 10 mins before being introduced into the defect and positioned to fill all void areas (Figure 5). Derived from bovine bone, the Endobon material has been used since 1989 as a bone replacement material in the fields of orthopedic and skull surgery but only recently introduced to the field of oral regeneration. Unlike most xenografts used in oral regenerative procedures, the bone is sintered, a process that involves heating it to more than 1200ºC. This removes all pathogenic and organic components from the bovine bone and yields a highly crystalline material containing small amounts of calcium oxide resulting from decomposition of the original carbon content. The sintered bovine bone incorporates macroporosities but preserves the microporosity of the original bone.

A dry 20 mm x 30 mm OsseoGuard® (cross-linked) Resorbable Collagen Membrane (BIOMET *3i*) was trimmed to size and placed under the facial flap prior to rehydration. The insertion of the dry membrane allowed the operator to easily verify that the membrane covered the apex of the lesion (Figure 6). It was allowed to hydrate with the patient's blood while under the gingival flap (Figure 7).

Extensive periosteal-releasing incisions were made in the facial flap to allow for passive primary closure. An Essix removable partial denture (DENTSPLY Raintree Essix, www.essix.com) was delivered, and it was verified that no contact was being made with the ridge. The patient was given

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prescriptions for antibiotics and analgesics. He returned for follow-up at 7 days, 14 days, 1 month, 3 months, and 6 months, respectively.

Six months postoperative, a crestal incision with distal release was made and a full-thickness flap was reflected, revealing excellent hard-tissue fill. A 2-mm trephine core was harvested from the facial aspect of the newly regenerated bone and stored in formalin until further histologic evaluation could be performed. Encode® Healing Abutments (BIOMET *3i*) were placed into the two internally interfaced implants (Figure 8), and primary closure obtained around the abutments. The Essix retainer was repositioned, and the patient was dismissed.

After 6 weeks of soft-tissue healing, the patient returned for restorative treatment. The definitive three-unit bridge with a cantilever at the right lateral incisor was fabricated and inserted. A radiograph taken at this appointment provided evidence that the crestal height, interproximal bone, and circumferential bone had been preserved, allowing for maintenance of the interproximal papillae (Figure 9 and Figure 10).

HISTOLOGIC PROCESSING AND EXAMINATION

The trephine core sample (6 mm in length and 2 mm in diameter) that was harvested at the time of implant uncovering was dehydrated in increasing grades of ethanol and subsequently infiltrated with Technovit® 7200 VLC-resin (Kulzer, www.kulzer-technik.de). Following embedding in the acrylic resin, the cylinder was polymerized and sectioned vertically using a cutting–grinding unit (Exakt®, Apparatebau GmbH, www.exaktusa.com). The 250-μm thick units obtained were further reduced by microgrinding and polishing to a final thickness of approximately 20 μm to 30 μm. The sections were stained with hematoxylin and eosin and examined under a Leitz DM-RBE® light microscope (Leica, www.leica-microsystems.com) equipped with an image system (Q-500 MC®, Leica).

The microscopic examination revealed a number of unresorbed particles, well embedded in the newly formed bone trabeculae. This finding is consistent with histologic examination of other commercially available xenografts (Figure 11 and Figure 12).

CONCLUSION

The 6-month clinical and histologic results obtained in this case suggest Endobon Xenograft Granules used with OsseoGuard

Resorbable Collagen Membranes are viable regenerative materials for cases in which intra-alveolar, space-maintaining deficiencies prevent proper implant placement. Combining these regenerative materials with a platform-switched implant proved to be a successful strategy for anterior esthetic rehabilitation. Further studies with larger patient populations are needed to confirm these results.

REFERENCES

1. Almer M, Johnson PL, Salman L. Histological and histochemical investigation of human alveolar socket healing in undisturbed extraction wound. *J Am Dent Assoc.* 1960;61:46-48.

2. Atwood DA. Reduction of residual ridges: a major oral disease entity. *J Prosthet Dent.* 1971;26(3):266-279.

3. Schropp L, Wenzel A, Kostopoulos L, et al. Bone healing and soft tissue contour changes following single-tooth extraction: a clinical and radiographic 12-month prospective study. *Int J Periodontics Restorative Dent.* 2003;23(4):313-323.

4. Cardaropoli G, Araújo M. Lindhe J. Dynamics of bone tissue formation in tooth extraction sites. An experimental study in dogs. *J Clin Periodontol.* 2003;30(9):809-818.

5. Araújo MG, Lindhe J. Dimensional ridge alterations following tooth extraction. An experimental study in the dog. *J Clin Periodontol.* 2005;32(2):212-218.

6. Lekovic V, Kenney EB, Weinlaender M, et al. A bone regenerative approach to alveolar ridge maintenance following tooth extraction. Report of 10 cases. *J Periodontol.* 1997;68(6):563-570.

7. Nevins M, Camelo M, De Paoli S, et al. A study of the fate of buccal wall of extraction sockets of teeth with prominent roots. *Int J Periodontics Restorative Dent.* 2006;26:19-29.

8. Fugazzoto PA. Treatment options following single-rooted tooth removal: a literature review and proposed hierarchy of treatment selection. *J Periodontol.* 2005;76(5):821-831.

9. Pietrokovski J, Massler M. Alveolar ridge resorption following tooth extraction. *J Prosthet Dent.* 1967;17(1):21-27.

10. Duncan JM, Westwood RM. Ridge widening for the thin maxilla: a clinical report. *Int J Oral Maxillofac Implants.* 1997;12(2): 224-227.

11. Scipioni A, Bruschi G, Calesini G. The edentulous ridge expansion technique: a five-year study. *Int J Periodontics Restorative Dent.* 1994;14(5):451-459.

12. Vercellotti T. The Piezoelectric surgery in implantology: a case report—a new piezoelectric ridge expansion technique. *Int J Periodontics Restorative Dent.* 2000;20:358-365.

13. Mellonig J, Nevins M. Guided bone regeneration of bone defects associated with implants: an evidence-based outcome assessment. *Int J Periodontics Restorative Dent.* 1995;15(2):168-185.

14. Zitzmann N, Naef R, Schärer P. Resorbable versus nonresorbable membranes in combination with Bio-Oss for guided bone regeneration. *Int J Oral Maxillofac Implants.* 1997;12(6):844-852.

15. Urbani G, Lombardo G, Santi E, et al. Distraction osteogenesis to achieve mandibular vertical bone regeneration: a case report. *Int J Periodontics Restorative Dent.* 1999;19(4):321-331.

16. Proussaefs P, Lozada J. The use of intraorally harvested autogenous block grafts for vertical alveolar ridge augmentation: a human study. *Int J Periodontics Restorative Dent.* 2005;25(4):351-363.

17. Misch CM. Comparison of intraoral donor sites for onlay grafting prior to implant placement. *Int J Oral Maxillofac Implants.* 1997;12(6):767-776.

18. Pikos MA. Block autografts for localized ridge augmentation: part I. The posterior maxilla. *Implant Dent.* 1999;8(3):279-285.

19. Jensen J, Reiche-Fischel O, Sindet-Pedersen S. Nerve transposition and implant placement in the atrophic posterior mandibular alveolar ridge. *J Oral Maxillofac Surg.* 1994;52(7):662-668.

20. Boyne PJ, James RA. Grafting of the maxillary sinus floor with autogenous marrow and bone. *J Oral Surg.* 1980;38(8):613-616.

21. Vercellotti T, De Paoli S, Nevins M. The Piezoelectric bony window osteotomy and sinus membrane elevation: introduction of a new technique for simplification of the sinus augmentation procedure. *Int J Periodontics Restorative Dent.* 2001;21(6):561-567.

22. Misch CM. Comparison of intraoral donor sites for onlay grafting prior to implant placement. *Int J Oral Maxillofac Implants.* 1997;12(6):767-776.

23. Pikos MA. Block autografts for localized ridge augmentation: part I. The posterior maxilla. *Implant Dent.* 1999;8(3):279-285.

24. Rudman RA. Prospective evaluation of morbidity associated with iliac crest harvest for alveolar cleft grafting. *J Oral Maxillofac Surg.* 1997;55(3):219-223.

25. Schwartz-Arad D, Levin L, Sigal L. Surgical success of intraoral autogenous block onlay grafting for alveolar ridge augmentation. *Implant Dent.* 2005;14(2):131-138.

26. Raghoebar G, Louwerse C, Kalk WW, et al. Morbidity of chin bone harvesting. *Clin Oral Implants Res.* 2001;12(5):503-507.

27. Clavero J, Lundgren S. Ramus or chin grafts for maxillary sinus inlay and local onlay augmentation: comparison of donor site morbidity and complications. *Clin Implant Dent Relat Res.* 2003;5(3):154-160.

28. Misch CM, Misch CE, Resnik RR, et al. Reconstruction of maxillary alveolar defects with mandibular symphysis grafts for dental implants: a preliminary procedural report. *Int J Oral Maxillofac Implants.* 1992;7(3):360-366.

29. Widmark G, Andersson B, Ivanoff C. Mandibular bone graft in the anterior maxilla for single-tooth implants. Presentation of surgical method. *Int J Oral Maxillofac Surg.* 1997;26(2):106-109.

30. McAllister B, Haghighat K. Bone augmentation techniques. *J Periodontol.* 2007;78(3):377-396.

31. Proussaefs P, Lozada J. The use of intraorally harvested autogenous block grafts for vertical alveolar ridge augmentation: a human study. *Int J Periodontics Restorative Dent.* 2005;25(4):351-363.

32. Aghaloo TL, Moy PK. Which hard tissue augmentation techniques are the most successful in furnishing bony support for implant placement? *Int J Oral Maxillofac Implants.* 2007;22(suppl):49-70.

33. Lupovici J. Revisiting the hopeless ridge: part I—challenging the gold standard. *Compend Contin Educ Dent.* 2009;30(3):130-140.

34. Becker W, Becker BE, Caffesse R. A comparison of demineralized freeze-dried bone and autogenous bone to induce bone formation in human extraction sockets. *J Periodontol.* 1994;65(12):1128-1133.

35. Becker W, Clokie C, Sennerby L, et al. Histologic findings after implantation and evaluation of different graft materials and titanium micro screws into extraction sockets: case reports. *J Periodontol.* 1998;69(4):414-421.

36. Schwartz Z, Mellonig JT, Carnes DL Jr, et al. Ability of commercial demineralized freeze-dried bone allograft to induce new bone formation. *J Periodontol.* 1996;67(9):918-926.

37. Artzi Z, Tal H, Dayan D. Porous bovine bone mineral in healing of human extraction sockets: 2. Histochemical observations at 9 months. *J Periodontol.* 2001;72(2):152-159.

38. Wenz B, Oesch B, Horst M. Analysis of the risk of transmitting bovine spongiform encephalopathy through bone grafts derived from bovine bone. *Biomaterials.* 2001;22(12):1599-1606.

39. Sogal A, Tofe AJ. Risk assessment of bovine spongiform encephalopathy transmission through bone graft material derived from bovine bone used for dental applications. *J Periodontol.* 1999;70(9):1053-1063.

40. Froum SJ, Wallace SS, Cho S-C, et al. Use of xenografts for sinus augmentation. In: Jensen OT, ed. *The Sinus Bone Graft.* 2nd ed. Chicago, IL: Quintessence; 2006:211-219.

41. Becker W, Clokie C, Sennerby L, et al. Histologic findings after implantation and evaluation of different grafting materials and titanium microscrews into extraction sockets: case reports. *J Periodontol.* 1998;69(4):414-421.

42. Pinholt EM, Ruyter IE, Haanaes HR, et al. Chemical, physical, and histologic studies on four commercial apatites used for alveolar ridge augmentation. *J Oral Maxillofac Surg.* 1992;50(8):859-867.

43. Peetz M. Characterization of xenogrenic bone material. In: Boyne PJ, Evensen L, eds. *Osseous Reconstruction of the Maxilla and the Mandible: Surgical Techniques Using Titanium Mesh and Bone Mineral.* Carol Stream, IL: Quintessence; 1997:87-100.

44. McAllister BS, Margolin MD, Cogan AG, et al. Residual lateral wall defects following sinus grafting with recombinant human osteogenic protein-1 or Bio-Oss in the chimpanzee. *Int J Periodontics Restorative Dent.* 1998;18(3):227-239.

45. Klinge B, Alberius P, Isaksson S, et al. Osseous response to implanted natural bone mineral and synthetic hydroxylapatite ceramics in the repair of experimental skull bone defects. *J Oral Maxillofac Surg.* 1992;50(3):241-249.

46. Spector M. Anorganic bovine bone and ceramic analogs of bone mineral as implants to facilitate bone regeneration. *Clin Plast Surg.* 1994;21(3):437-444.

47. Hämmerle CH, Olah AJ, Schmid J, et al. The biological effect of natural bone mineral on bone neoformation on the rabbit skull. *Clin Oral Implants Res.* 1997;8(3):198-207.

48. Hämmerle CH, Chiantella Gc, Karring T, et al. The effect of a deproteinized bovine bone mineral on bone regeneration around titanium dental implants. *Clin Oral Implants Res.* 1998;9 (3):151-162.

49. Jensen SS, Aaboe M, Pinholt EM, et al. Tissue reaction and material characteristics of four bone substitutes. *Int J Oral Maxillofac Implants.* 1996;11(1):55-66.

50. Jovanovic S, Nevins M. Bone formation utilizing titanium-reinforced barrier membranes. *Int J Periodontics Restorative Dent.* 1995;15(1):56-69.

51. von Arx T, Hardt N, Wallkamm B. The TIME technique: a new method for localized alveolar ridge augmentation prior to placement of dental implants. *Int J Oral Maxillofac Implants.* 1996;11(3):387-394.

52. Machtei EE. The effect of membrane exposure on the outcome of regenerative procedures in humans: a meta-analysis. *J Periodontol.* 2001;72(4):512-516.

53. Simion M, Baldoni M, Rossi P, et al. A comparative study of the effectiveness of e-PTFE membranes with and without early exposure during the healing period. *Int J Periodontics Restorative Dent.* 1994;14(2):166-180.

54. Quteish D, Singrao S, Dolby AE. Light and electron microscopic evaluation of biocompatibility, resorption, and penetration characteristics of human collagen graft material. *J Clin Periodontol.* 1991;18(5):305-311.

55. Sableman E. Biology, biotechnology and biocompatability of collagen. In: William DF, ed. *Biocompatability of Tissue Analogs.* 1st ed. Boca Raton, FL: CRC Press; 1985:27.

56. Postlethwaite AE, Seyer JM, Kang AH. Chemotactic attraction of human fibroblasts to type I, II, III collagens and collagen-derived peptides. *Proc Natl Acad Sci U S A.* 1978;75(2):871-875.

57. Yaffe A, Ehrlich J, Shoshan A restoration of periodontal attachment employing collagen solution in the dog. *J Periodontal.* 1984;55(11):623-628.

58. Marinucci L, Lilli C, Baroni T, et al. In vitro comparison of bioabsorbable and non-resorbable membranes in bone regeneration. *J Periodontol.* 2001;72(6):753-759.

59. Rothamel D, Schwarz F, Sager M, et al. Biodegradation of different cross-linked membranes: an experimental study in the rat. *Clin Oral Implants Res.* 2005;16(3):369-378.

60. Bunyaratavej P, Wang HL. Collagen membranes: a review. *J Periodontol.* 2001;72(2):215-229.

61. Bornstein M, Bosshardt D, Buser D, et al. Effect of two different bioabsorbable collagen membranes on guided bone regeneration: A comparative histomorphometric study in the dog mandible. *J Periodontol.* 2007;78(10):1943-1953.

62. Freidmann A, Strietzel FP, Maretzki B, et al. Observations on a new collagen barrier membrane in 16 consecutively treated patients. Clinical and histological findings. *J Periodontal.* 2001;72(11):1616-1623.

63. Albrektsson T, Zarb G, Worthington P, et al. The long-term efficacy of currently used dental implants: a review and proposed criteria of success. *Int J Oral Maxillofac Implants.* 1986;1(1):11-25.

64. Cappiello M, Luongo R, Di Iorio D, et al. Evaluation of peri-implant bone loss around platform-switched implants. *Int J Periodontics Restorative Dent.* 2008;28(4):347-355.

65. Calvo Guirado JL, Saez Yuguero MR, Pardo Zamora G, et al. Immediate provisionalization on a new implant design for esthetic restoration and preserving crestal bone. *Implant Dent.* 2007;16(2):155-164.

66. Vela-Nebot X, Rodríguez-Ciurana X, Rodado-Alonso C, et al. Benefits of an implant platform modification technique to reduce crestal bone resorption. *Implant Dent.* 2006;15(3):313-320.

67. Levin L, Herzberg R, Dolev E, et al. Smoking and complications of onlay bone grafts and sinus lift operations. *Int J Oral Maxillofac Implant.* 2004;19(3):369-373.